

Molecular dissection of the GOLDEN1-LIKE gene family in rice

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Abstract

Drought stress has become one of the most severe bottlenecks experienced in rice production across the world. Plant responses to drought involve stomatal movement in the leaves, which is mainly regulated by abscisic acid (ABA). A previous study from our lab showed that GOLDEN1-LIKE (GL) genes are involved in stomatal movement in rice. In this study, we improved the function of ZmGL1 in rice and found that it can improve stomatal conductance and photosynthesis under field conditions. In the present study, we uncovered the function of ZmGL1 in rice in response to drought stress. We found that the elevated drought tolerance in rice plants overexpressing ZmGL1 or ZmGL2 is conferred by rapid ABA-mediated stomatal closure. Comparative analysis of RNA-sequence, RNA-seq data from the rice leaves and DNA microarray data on sequencing, DAP-seq results obtained in vivo revealed that ZmGL1 played roles in regulating ABA-related drought stress-responsive genes. Four upstream regulatory genes closely functioning in drought stress tolerance, including bHLH1, bHLH2, bHLH3 and bHLH4, were identified as putative regulatory genes of ZmGL1 and ZmGL2 in rice. These results demonstrate that ZmGL1 plays an important role in regulating stomatal movement and photosynthesis and stress tolerance. This study provides a new strategy for breeding drought-tolerant crop plants without compromising photosynthesis.

de'c mor re exposed o'oneg env ronmen'ls mul'p' b' h'm'ous m' b' r' hum' d' m' h' CO₂ levels, and p' h'o- y'ens' s'om' m' re r' p'oly closed, espec'ly p' d' osperms Serl' e' m' 2018' The s' dyn' m' c' movemen' s' driven by m' or pressure ch' d' es n' h' e' u' r' d' cells, s' resu' m' t' h' e' c' m' an of d' on ch' d' n' e' b' d' n' h' b' an of n' r' d' rec' d' n' y' + ch' d' n' e' b' h' ch' encode' g' by

pl' y' enes m' e' m' 2010' The g' flux of d' ions and sm' ll m' e' m' o' l' os, n' cl' u' d' i' n' g' C' F₃, NO₃⁻, and m' d' u' c' t' u' s' m' e' m' b' r' u' e' d' e' p' o' l' i' z' a' t' i' o' n' o' c' o' n' s' e' q' u' e' n' c' e' o' u' t' r' a' c' t' i' o' n' s' + ch' d' n' e' b' d' n' e' f' c' i' o' n' s' + e' f' f' l' u' x, f' u' r' t' h' e' r' r' e' d' u' c' i' n' g' m' o' r' e' or p' r' e' s' s' u' r' e' n' s' i' g' n' i' c' a' n' t' l' y' u' r' d' cells and l' e' d' i' n' g' o' t' h' e' s' o' m' e' c' l' o' s' u' r' e' P' u' d' e' y' e' m' 2007' Under t' h' e' d' e' c' c' o' n' d' i' t' i' o' n' s, t' h' e' p' h' y' t' o' r' m' o' n' e' a' b' s' c' i' s' c' i' c' ABA' p' l' y' s' s' h' e' p' r' i' m' a' r' y' r' e' s' u' l' t' o' f' s' o' m' e' c' l' o' s' u' r' e' m' o' v' e' m' e' n' t' p' r' e' v' e' n' t' i' o' n' o' f' s' s, n' t' h' e' e' n' d' o' g' y' e' n' o' u' s' ABA' l' e' v' e' l' s' o' r' e' c' o' n' t' r' o' l' l' e' d' b' y' p' r' e' c' i' s' e' b' a' l' a' n' c' e' b' e' t' w' e' e' n' b' o' s' y' n' t' h' e' s' i' s' and c' l' o' s' u' r' e, t' h' e' t' o' o' i' n' f' l' u' e' n' c' e' d' b' y' m' i' t' o' s' p' o' r' o' u' s' c' o' n' j' u' n' c' t' i' o' n' p' r' o' c' e' s' s' u' s' t' r' o' e' m' 2007' b' y' e' m' 2011' ABA' s' i' n' t' e' m' p' t' l' y' s' y' n' t' h' e' s' e' d' f' r' o' m' C₃ c' o' r' r' o' s' i' o' n' s' o' f' o' r' m' x' a' n' t' h' o' p' h' y' l' l' s' e' x' p' r' e' s' s' i' o' n, s' f' o' r' m' e' d' i' n' t' h' e' p' l' a' s' t' i' d' s' v' i' o' x' o' d' i' z' a' t' i' o' n' s' e' c' o' n' t' r' o' l' l' e' d' b' y' 9' -e' p' o' x' y' c' o' r' r' o' s' i' o' n' o' f' d' o' x' y' e' n' z' y' m' e' NCEP' d' u' o' x' i' n' s' i' n' e' n' e' x' p' o' r' t' e' d' o' t' h' e' c' y' t' o' s' o' l' and c' o' n' v' e' r' t' e' d' o' ABA' d' r' o' u' g' h' t' r' e' s' s' r' e' c' o' m' p' e' n' s' a' t' i' o' n' v' i' o' s' t' o' r' o' c' h' i' n' d' e' f' y' a' r' o' u' n' d' e' n' s' e' / r' e' d' u' c' t' i' o' n' 1' SDR1/ABA' d' Ar' h' o' p' o' s' i' s' i' n' t' e' r' y' a' e' o' x' i' d' a' s' e' 3' AAO3' Seo' and o' s' t' b' 2007; > o' n' y' and Z' h' u' 2003'

Tr' a' n' s' c' r' i' p' t' i' o' n' f' a' c' t' o' r' s' TFs' r' e' c' r' u' c' i' a' l' r' e' g' u' l' a' t' o' r' s' o' f' m' a' n' y' b' o' t' a' n' i' c' a' l' p' r' o' c' e' s' s' e' s, i' n' c' l' u' d' i' n' g' r' e' s' p' o' n' s' e' s' o' f' e' n' v' i' r' o' n' m' e' n' t' a' l' s' t' r' e' s' s' and h' o' r' m' o' n' e' r' e' g' u' l' a' t' i' o' n' . T' h' e' s' e' r' e' g' u' l' a' t' o' r' y' f' u' n' c' t' i' o' n' s' r' e' c' o' m' p' l' e' t' e' d' d' r' o' u' g' h' t' b' i' n' d' i' n' g' o' f' s' p' e' c' i' f' i' c' o' l' e' m' e' n' t' a' n' i' n' t' h' e' p' r' o' m' o' t' o' r' r' e' g' i' o' n' s' o' f' m' a' n' y' e' n' z' y' m' e' s' T' o' d' a' y' e' m' 2010' N' u' m' e' r' o' u' s' b' o' t' a' n' i' c' s' t' r' e' s' s' - r' e' s' p' o' n' s' e' v' e' TFs' h' a' v' e' b' e' e' n' d' e' n' t' i' f' i' e' d' i' n' p' l' a' n' t' s' f' o' r' i' n' s' t' a' n' c' e' s' WR' Y, MYB, and DREB/ CBF TFs' h' a' v' e' b' e' e' n' r' e' p' o' r' t' e' d' a' s' k' e' y' r' e' g' u' l' a' t' o' r' s' o' f' p' l' a' n' t' s' t' r' e' s' s' r' e' s' p' o' n' s' e' s' M' u' n' e' m' 2011' G' O' L' D' E' N' O' - I' I' E' G' L' f' l' TFs' e' n' e' r' g' i' c' a' l' t' r' a' n' s' c' r' i' p' t' i' o' n' a' l' c' o' n' t' r' o' l' l' e' r' s' o' f' c' h' l' o' r' o' p' h' y' l' l' d' e' v' e' l' o' p' m' e' n' t' and b' o' t' a' n' i' c' s' Ross' n' e' m' 2001; W' u' e' m' 2013' and p' l' a' n' t' i' m' p' o' r' t' a' n' t' o' l' e' r' a' n' c' e' i' n' r' e' g' u' l' a' t' o' r' y' n' u' c' l' e' a' r' p' h' o' s' y' n' t' h' e' s' i' s' - r' e' l' a' t' e' d' y' e' n' e' s' Chen' e' m' 2016' I' n' m' a' n' y' e' m' 2010' y' e' n' e' s,

h' a' v' e' s' h' o' w' n' d' i' f' f' e' r' e' n' t' i' a' l' e' x' p' r' e' s' s' i' o' n' p' a' t' t' e' r' n' s' b' e' t' w' e' e' n' m' e' s' o' p' h' y' l' l' c' e' l' l' s' and t' h' e' b' u' n' d' l' e' s' H' u' i' e' m' 1998; C' h' u' e' m' 2012' E' c' o' n' o' m' i' c' o' v' e' r' e' x' p' r' e' s' s' i' o' n' o' f' m' a' n' y' e' n' z' y' m' e' s' i' n' r' e' c' o' n' d' u' c' t' i' o' n' i' n' d' u' c' e' s' c' h' l' o' r' o' p' h' y' l' l' d' e' v' e' l' o' p' m' e' n' t' i' n' b' u' n' d' l' e' s' h' e' t' i' c' c' e' l' l' s' and c' o' n' t' r' o' l' l' e' s' i' n' t' e' l' l' u' l' a' r' p' l' a' s' t' i' d' e' s' m' a' t' u' r' e' c' o' n' n' e' c' t' i' o' n' s, c' o' n' s' i' d' e' r' i' n' g' t' h' e' k' e' y' s' t' e' p' i' n' f' o' r' m' i' n' g' n' o' r' m' a' l' p' r' o' t' e' i' n' s' i' n' t' h' e' m' a' t' u' r' e' m' o' n' f' r' o' m' C₃ o' C₄ p' h' o' s' y' n' t' h' e' s' i' s' W' u' e' m' 2017' A' p' r' e' v' i' o' u' s' s' t' u' d' y' f' r' o' m' o' u' r' l' a' b' s' h' o' w' e' d' t' h' a' t' c' o' n' s' t' a' n' t' e' x' p' r' e' s' s' i' o' n' i' n' r' e' c' e' l' l' s' i' n' c' r' e' a' s' e' d' x' a' n' t' h' o' p' h' y' l' l' c' o' n' t' e' n' t' and f' u' r' t' h' e' r' m' a' t' u' r' e' s' t' h' e' p' h' o' s' y' n' t' h' e' s' i' s' u' n' d' e' r' h' y' h' - l' y' t' i' c' o' n' d' i' t' i' o' n' s, r' e' s' u' l' t' i' n' g' i' n' a' n' e' i' g' h' t' e' d' p' h' o' s' y' n' t' h' e' s' i' s' p' r' o' d' u' c' t' i' o' n' o' f' h' y' t' h' e' r' s' o' m' e' c' l' o' s' u' r' e' d' u' c' t' i' o' n' and i' m' p' r' o' v' e' d' b' i' o' m' a' s' s' and y' i' e' l' d' i' n' t' h' e' f' i' e' l' d' L' e' e' m' 2010' M' o' r' e' o' v' e' r, G' L' s' t' o' f' u' n' c' t' i' o' n' i' n' b' o' t' a' n' i' c' s' t' r' e' s' s' r' e' s' p' o' n' s' e' s' A' h' m' a' d' e' m' 2019' and p' h' o' s' y' n' t' h' e' s' i' s' r' e' s' p' o' n' s' e' s'

Murmu' e' m' 2014' f' o' r' e' x' a' m' p' l' e, G' L' s' i' n' c' r' e' a' s' e' s' m' a' t' u' r' e' m' o' v' e' m' e' n' t' i' n' A' r' b' i' d' o' p' s' i' n' t' h' e' n' e' x' p' o' s' e' d' o' o' n' g' e' N' u' s' h' e' m' 2016'

I' n' t' h' i' s' s' t' u' d' y, t' h' e' u' n' c' o' v' e' r' e' d' t' h' e' d' u' a' l' f' u' n' c' t' i' o' n' o' f' m' a' n' y' G' L' s, and t' h' e' e' c' c' e' n' t' r' i' c' o' v' e' r' e' x' p' r' e' s' s' i' o' n' o' f' t' h' e' i' n' t' e' n' s' i' t' y' i' n' r' e' c' e' n' t' e' r' e' d' i' m' p' r' o' v' e' d' d' r' o' u' g' h' t' t' o' l' e' r' a' n' c' e' b' y' p' r' o' m' o' t' i' n' g' s' o' m' e' c' l' o' s' u' r' e' i' n' r' e' s' p' o' n' s' e' o' t' h' e' r' d' e' c' c' o' n' t' r' o' l' l' e' m' a' t' u' r' e' i' n' t' h' e' s' o' m' e' c' l' o' s' u' r' e' c' o' n' d' u' c' t' i' o' n' o' b' o' t' a' n' i' c' e' n' t' p' h' o' s' y' n' t' h' e' s' i' s' i' n' s' u' f' f' i' c' i' e' n' t' a' n' d' s' u' s' t' a' i' n' a' b' l' e' W' e' f' u' r' t' h' e' r' s' h' o' w' e' d' t' h' a' t' p' l' a' n' t' s' o' m' e' c' l' o' s' u' r' e' m' o' v' e' m' e' n' t' o' s' m' e' d' i' a' t' e' d' b' y' ABA- i' n' v' o' l' v' e' d' p' r' o' t' e' i' n' s' u' n' d' e' r' d' r' o' u' g' h' t' c' o' n' d' i' t' i' o' n' s. T' h' e' s' e' r' e' s' u' l' t' s' s' u' g' g' e' s' t' t' h' a' t' y' e' n' e' s' m' a' y' b' e' p' r' o' m' o' t' i' n' g' c' a' n' d' i' d' a' t' e' s' f' o' r' b' r' e' e' d' i' n' g' r' e' c' o' v' e' r' e' s' o' f' h' y' t' h' e' r' s' o' m' e' c' l' o' s' u' r' e' f' l' e' x' i' b' i' l' i' t' y' and s' u' s' t' a' i' n' a' b' l' e' y' e' l' d' i' n' t' h' e' f' i' e' l' d' s' o' n' l' y' i' m' p' r' o' v' e' s' r' e' c' u' l' a' r' p' r' o' d' u' c' t' i' o' n' and i' n' c' r' e' a' s' e' f' o' o' d' s' e' c' u' r' i' t' y' i' n' t' h' e' c' o' n' t' r' o' l' c' l' i' m' a' t' e' c' h' a' n' g' e' .

Results

ZmGL1 and ZmGL2 confer drought tolerance

I' n' o' u' r' p' r' e' v' i' o' u' s' s' t' u' d' y, ZmGL1 i' n' t' r' a' n' s' g' e' n' e' r' i' c' r' e' c' e' i' n' s' c' o' n' s' t' a' n' t' l' y' e' x' p' r' e' s' s' i' n' g' o' r' i' n' d' u' c' e' d' b' y' t' h' e' m' a' t' u' r' e' o' f' p' r' o' m' o' t' o' r' p' e' r' f' o' r' m' e' d' i' m' p' r' o' v' e' d' p' h' o' s' y' n' t' h' e' s' i' s' r' a' t' e' s' and h' y' t' h' e' r' s' o' m' e' c' l' o' s' u' r' e' L' e' e' m' 2010' W' e' f' u' r' t' h' e' r' e' x' p' r' e' s' s' e' d' t' h' e' s' o' m' e' c' l' o' s' u' r' e' r' e' s' p' o' n' s' e' s' o' f' m' a' t' u' r' e' i' n' t' r' a' n' s' g' e' n' e' r' i' c' r' e' c' e' i' n' t' h' e' d' e' c' c' o' n' t' r' o' l' l' e' d' e' x' p' e' r' i' m' e' n' t' i' n' t' h' e' r' o' t' a' t' i' o' n' . S' u' r' p' r' i' s' i' n' g' l' y, m' a' t' u' r' e' i' n' t' r' a' n' s' g' e' n' e' r' i' c' r' e' c' e' i' n' t' h' e' s' o' m' e' c' l' o' s' u' r' e' d' r' o' u' g' h' t' t' o' l' e' r' a' n' c' e' i' n' t' h' e' t' e' s' t' o' p' e' W' T' p' l' a' n' t' s' f' u' l' l' r' e' c' o' v' e' r' y' f' r' o' m' 10- d' a' t' d' r' o' u' g' h' t' t' o' r' e' c' o' v' e' r' e' d' F₂ 1A' S' p' e' c' i' f' i' c' a' l' l' y, t' h' e' s' u' r' v' i' v' a' l' r' a' t' e' s' o' f' : : : o' p' l' a' n' t' s' w' e' r' e' 53.0% o' 64.0% f' o' r' t' h' e' 6- d' r' e' c' o' v' e' r' y' p' e' r' i' o' d, t' h' e' r' e' s' t' i' n' t' e' c' o' n' t' r' o' l' l' e' d' h' y' t' h' e' r' i' n' t' h' e' W' T' 14.9%; F₂ 1B' M' o' r' e' o' v' e' r, t' h' e' r' e' l' a' t' i' v' e' w' a' t' e' r' c' o' n' t' e' n' t' (RWC) i' n' t' h' e' l' e' a' v' e' s' o' f' W' T' and m' a' t' u' r' e' i' n' t' r' a' n' s' g' e' n' e' r' i' c' r' e' c' e' i' n' t' h' e' f' i' e' l' d' f' r' o' m' 94.7% o' 95.3% b' e' f' o' r' e' d' r' o' u' g' h' t' t' o' d' e' c' r' e' a' s' e' d' o' 73.1% i' n' t' h' e' W' T' f' o' r' t' h' e' t' e' s' t' o' p' e' i' n' t' h' e' f' i' e' l' d' f' o' r' 7 d' i' n' c' o' m' p' a' r' i' s' o' n, : : : o' p' l' a' n' t' s' i' n' t' h' e' t' e' s' t' o' p' e' i' n' t' r' a' n' s' g' e' n' e' r' i' c' r' e' c' e' i' n' t' h' e' f' i' e' l' d' f' o' r' 7 d' i' n' c' o' m' p' a' r' i' s' o' n, : : : p' l' a' n' t' s' i' n' t' h' e' f' i' e' l' d' r' e' l' a' t' i' v' e' l' y' h' y' t' h' e' r' RWC, e' s' p' e' c' i' a' l' l' y : : : r' e' c' o' v' e' r' y' f' r' o' m' 86.7% o' 90.9% A' f' t' e' r' 10 d' o' f' d' r' o' u' g' h' t' s' t' r' e' s' s, t' h' e' RWC' v' a' l' u' e' s' o' f' W' T' and : : : p' l' a' n' t' s' d' e' c' r' e' a' s' e' d' o' 11.6% o' 17.9%, t' h' e' r' e' s' t' i' n' t' e' c' o' n' t' r' o' l' l' e' d' h' y' t' h' e' r' i' n' t' h' e' t' e' s' t' o' p' e' o' f' : : : o' p' l' a' n' t' s' o' 14.5% o' 18.6%; F₂ 1C' T' h' e' s' e' r' e' s' u' l' t' s' i' n' d' i' c' a' t' e' t' h' a' t' ZmGL1 and ZmGL2 b' o' t' h' c' o' n' f' e' r' r' e' d' h' y' t' h' e' r' c' o' n' t' r' o' l' l' e' d' e' x' p' r' e' s' s' i' o' n' and t' h' u' s' d' r' o' u' g' h' t' t' o' l' e' r' a' n' c' e' .

W' e' n' e' x' t' e' x' p' r' e' s' s' e' d' t' h' e' r' o' t' a' t' i' o' n' p' e' r' f' o' r' m' a' n' c' e' o' f' W' T, : : : : : : : : : r' e' c' e' p' t' i' v' e' PEG- i' n' d' u' c' e' d' o' s' m' o' t' i' c' s' t' r' e' s' s' s' u' s' t' a' i' n' a' b' l' e' m' a' t' u' r' e' m' o' n' . A' f' t' e' r' 7 r' o' t' a' t' i' o' n' s' o' f' PEG 6000 f' o' r' 10 d' o' : : : o' p' l' a' n' t' s' : : : o' r' e' c' e' p' t' i' v' e' s' h' o' w' e' d' l' e' s' s' i' n' j' u' r' y' and c' h' l' o' r' o' s' i' s' c' o' m' p' a' r' e' d' o' t' h' e' W' T' S' u' p' p' l' e' m' e' n' t' F₂ 51A' T' h' e' m' a' x' i' m' u' m' q' u' a' n' t' i' t' y' o' f' P' S' I' i' n' t' h' e' s' u' r' f' a' c' e' o' f' p' l' a' n' t' s' i' m' p' o' r' t' a' n' t' c' o' n' t' r' o' l' l' e' d' o' f' p' l' a' n' t' p' h' y' s' i' o' l' o' g' y' c' h' l' o' r' o' s' i' s' u' n' d' e' r' s' t' r' e' s' s' c' o' n' d' i' t' i' o' n' s, and t' h' e' t' e' s' t' o' p' e' .

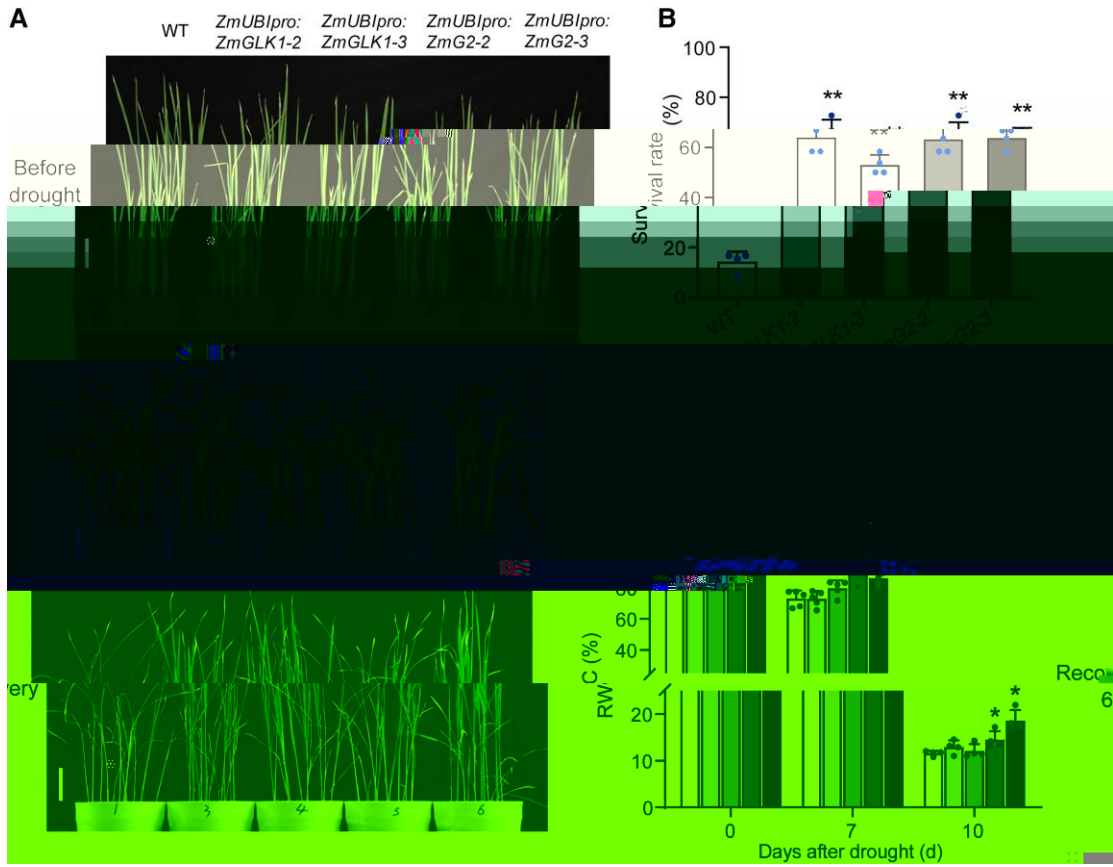


Figure 1. Overexpression of *ZmGLK1-3* and *ZmG2-3* in rice increases drought tolerance. **A)** Phenotypes of WT, *ZmGLK1-2*, *ZmGLK1-3*, *ZmG2-2*, and *ZmG2-3* rice plants during drought stress. Three-week-old WT, *ZmGLK1-2*, *ZmGLK1-3*, *ZmG2-2*, and *ZmG2-3* rice seedlings were grown in soil under drought stress by withholding water for 10 d and then rewatered for 6-d recovery period. The upper, middle, and lower panels show representative plants before drought stress, after 10 d of drought stress, and after the 6-d recovery, respectively. Scale bar = 2 cm. **B)** Survival rates of WT, *ZmGLK1-2*, *ZmGLK1-3*, *ZmG2-2*, and *ZmG2-3* rice plants after 10 d of drought stress followed by 6 d of recovery. Data are presented as the mean \pm SD from triplicate replicates. **C)** The RWC of WT, *ZmGLK1-2*, *ZmGLK1-3*, *ZmG2-2*, and *ZmG2-3* rice leaves after 0, 7, and 10 d of drought stress. Data are presented as the mean \pm SD.

10 d of PEG treatment. **Supplement F₇. S1B.** We also monitored changes of RWC in rice seedlings during PEG treatment. The results showed that the transgenic plants maintained significantly higher RWC compared to the WT. Specifically, RWC values were 11.4%, 10.1% and 9.5% and 9.7% higher in *ZmGLK1-2*, *ZmGLK1-3*, and *ZmG2-2*, respectively, compared to the WT. **Supplement F₇. S1C.** These results were further confirmed by overexpression of *ZmGLK1-3* and *ZmG2-3* in rice. Significantly improved drought and osmotic stress.

ZmGLK1 and ZmG2 regulate stomatal closure during drought stress

To further investigate the physiological mechanism underlying the elevated drought tolerance conferred by *ZmGLK1* and *ZmG2*, we evaluated the effect of drought stress on stomatal closure of rice seedlings in the pot in the field number, since stomatal closure is a key factor for water exchange and transpiration in plants serving as the dominant mechanism of photosynthesis under drought. We therefore measured

stomatal conductance and photosynthesis-related parameters under control conditions using a LiCOR-6400 Transpiration photosynthesis system. The results revealed significantly higher stomatal conductance in *ZmGLK1-2* and *ZmGLK1-3* rice seedlings (0.118–0.139 and 0.126–0.131, respectively) compared to the WT (0.083) under control conditions; the transgenic plants also performed higher photosynthesis rates, net cellular CO₂ concentrations (C_i) and transpiration rates. **Supplement F₇. S2.** In the pot, stomatal conductance in the field decreased significantly after 7 d of drought treatment in control and transgenic rice plants. *ZmGLK1-2* and *ZmGLK1-3* rice plants displayed significantly decrease in stomatal conductance (0.067–0.073 and 0.054–0.059, respectively) here, whereas the WT remained relatively stable under drought conditions (0.087). **Supplement F₇. S2B.** The photosynthesis rates, C_i, and transpiration rates showed correspondingly declines in *ZmGLK1-2* and *ZmGLK1-3* rice plants during drought deprivation. **Supplement F₇. S2, A, C, and D.** We next compared the stomatal conductance between WT and *ZmGLK1-2* or *ZmGLK1-3* rice plants under

both control and drought conditions. Transgenic plants represented higher stomatal density in the leaves but did not significantly show or stomatal conductance of the WT regardless of conditions (Fig. 2). Additionally, the stomata were prominently derived from the leaf epidermal cells under control conditions (Fig. 2D). Here, under drought stress, the stomatal density was significantly decreased in transgenic plants to a lower level than WT, consistent with the stomatal pore area (Fig. 2E).

Considering the relationship between stomatal density and chamber conductance, the further conductance experiments in the greenhouse were performed to exclude the influence of stomatal density. As expected, the results

showed consistency with the chamber experiment (Fig. 1). All plants were severely impacted due to the rapid loss of water during the 10-d drought duration (Supplemental Fig. S3; Fig. 3A). After re-watering for 7 d, the observed higher survival rate in the control plants (Fig. 3B) suggested that the stomatal density higher RWC of the leaves in WT after during the drought may be recovery (Fig. 3C). Moreover, the monitored dynamics of photosynthesis rate and stomatal conductance throughout the duration of drought and recovery (Fig. 3D): the control plants performed higher photosynthesis rate and stomatal conductance under sufficient water condition. Nevertheless, the photosynthesis rate (Fig. 3E) and

through a deepened, of h ch : and
receptor presence in photosynthesis
r. and the sum of conductance compared to the WT.
Fig. 3, D and E. These results clearly indicate

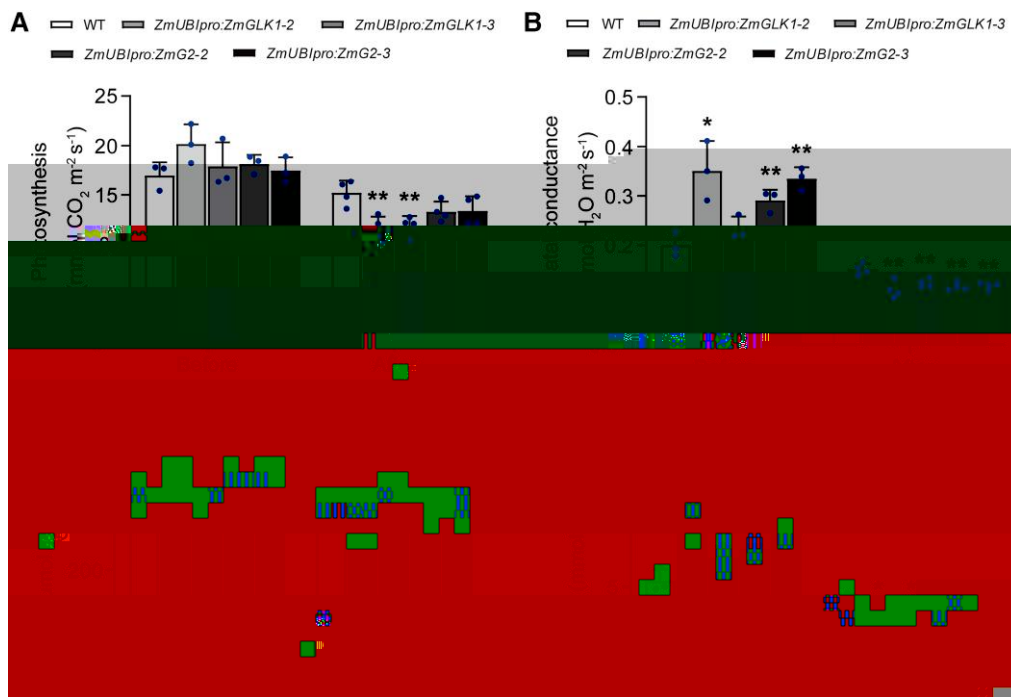


Figure 4. Exogenous ABA application reduced the photosynthesis rate and stomatal conductance in rice plants overexpressing or complementing the WT. **A)** Photosynthesis rates, **B)** stomatal conductance, **C)** C_i and **D)** transpiration rates of 3-year-old WT, ZmUBIpro:ZmGLK1-2, ZmUBIpro:ZmGLK1-3, ZmUBIpro:ZmG2-2, and ZmUBIpro:ZmG2-3 rice plants grown in soil before or for 5 h after ABA treatment. D values represent the mean \pm SD from 3 biological replicates. * < 0.05 , ** < 0.01 . Scale bars are as indicated.

The WT and transgenic plants mimicking the regulatory network from the drought stress response in rice plants showed a reduction of stomatal closure in response to exogenous stress conferred by ZmGL1 and ZmG2 to ABA mediated.

ZmGL1 and ZmG2 regulate drought tolerance

To further understand the molecular mechanisms regulated by ZmGL1s under drought stress, we next compared the expression levels of several key genes associated with stomatal movement in WT, ZmUBIpro:ZmGLK1-2, ZmUBIpro:ZmGLK1-3, ZmUBIpro:ZmG2-2, and ZmUBIpro:ZmG2-3 rice plants under control and drought stress conditions. Under control conditions, several key genes were highly expressed in the transgenic plants compared to the WT but profoundly downregulated in response to drought stress. These comprised genes encoding proteins associated with stomatal regulation such as *OST1*, *SLAC1*, *OST2*, *OST3*, *OST4*, *OST5*, *OST6*, *OST7*, *OST8*, *OST9*, *OST10*, *OST11*, *OST12*, *OST13*, *OST14*, *OST15*, *OST16*, *OST17*, *OST18*, *OST19*, *OST20*, *OST21*, *OST22*, *OST23*, *OST24*, *OST25*, *OST26*, *OST27*, *OST28*, *OST29*, *OST30*, *OST31*, *OST32*, *OST33*, *OST34*, *OST35*, *OST36*, *OST37*, *OST38*, *OST39*, *OST40*, *OST41*, *OST42*, *OST43*, *OST44*, *OST45*, *OST46*, *OST47*, *OST48*, *OST49*, *OST50*, *OST51*, *OST52*, *OST53*, *OST54*, *OST55*, *OST56*, *OST57*, *OST58*, *OST59*, *OST60*, *OST61*, *OST62*, *OST63*, *OST64*, *OST65*, *OST66*, *OST67*, *OST68*, *OST69*, *OST70*, *OST71*, *OST72*, *OST73*, *OST74*, *OST75*, *OST76*, *OST77*, *OST78*, *OST79*, *OST80*, *OST81*, *OST82*, *OST83*, *OST84*, *OST85*, *OST86*, *OST87*, *OST88*, *OST89*, *OST90*, *OST91*, *OST92*, *OST93*, *OST94*, *OST95*, *OST96*, *OST97*, *OST98*, *OST99*, *OST100*. These results demonstrated that ZmGL1 and ZmG2 improved drought tolerance by downregulating genes involved in stomatal movement when suffering from drought stress. A genome-wide transcriptomic analysis also conducted in WT, ZmUBIpro:ZmGLK1-2, ZmUBIpro:ZmGLK1-3, ZmUBIpro:ZmG2-2, and ZmUBIpro:ZmG2-3 rice plants 3 h after ABA treatment revealed that the effective ZmGL1 and ZmG2 introduced by ABA, especially

on stomatal movement. WT plants clearly showed a significant expression patterns compared to the transgenic plants. This analysis demonstrated by the cluster separation analysis the principal component analysis (PCA) (Fig. 6A). Specifically, for ABA treatment ZmUBIpro:ZmGLK1-2 and ZmUBIpro:ZmG2-2 rice plants upregulated 775 genes respectively. ZmUBIpro:ZmGLK1-3 and ZmUBIpro:ZmG2-3 plants respectively, compared to the WT, of which 485 genes were upregulated in both transgenic lines (Fig. 6B). Gene Ontology (GO) term enrichment analysis revealed that the upregulated differentially expressed genes (DEGs) in ZmUBIpro:ZmGLK1-2 and ZmUBIpro:ZmG2-2 plants functioned in multiple biological processes but primarily in the ABA signaling pathway (Fig. 6, C and D). Next, we performed DNA microarray sequencing (DAP-seq) analysis to identify genes regulated by the ZmGL1 TFs. This analysis revealed 6,601 and 6,565 putative binding sites of ZmGL1 and ZmG2 in the rice genome, respectively, and more than half of the identified sites being bound by both ZmGL1 and ZmG2 (Supplement Fig. S4A). Of the 3,835 binding sites shared by ZmGL1 and ZmG2, 17.5% were located in promoters, 8.59% exons, and 3.6% introns (Supplement Fig. S4B). Moreover, this analysis demonstrated that the most enriched motifs found in the ZmGL1- and ZmG2-binding regions were GCCTCT and AGATTCT (Supplement Fig. S4C and D). Furthermore, genes identified from the DAP-seq data were also identified from the RNA-sequencing (RNA-seq) data of differentially expressed

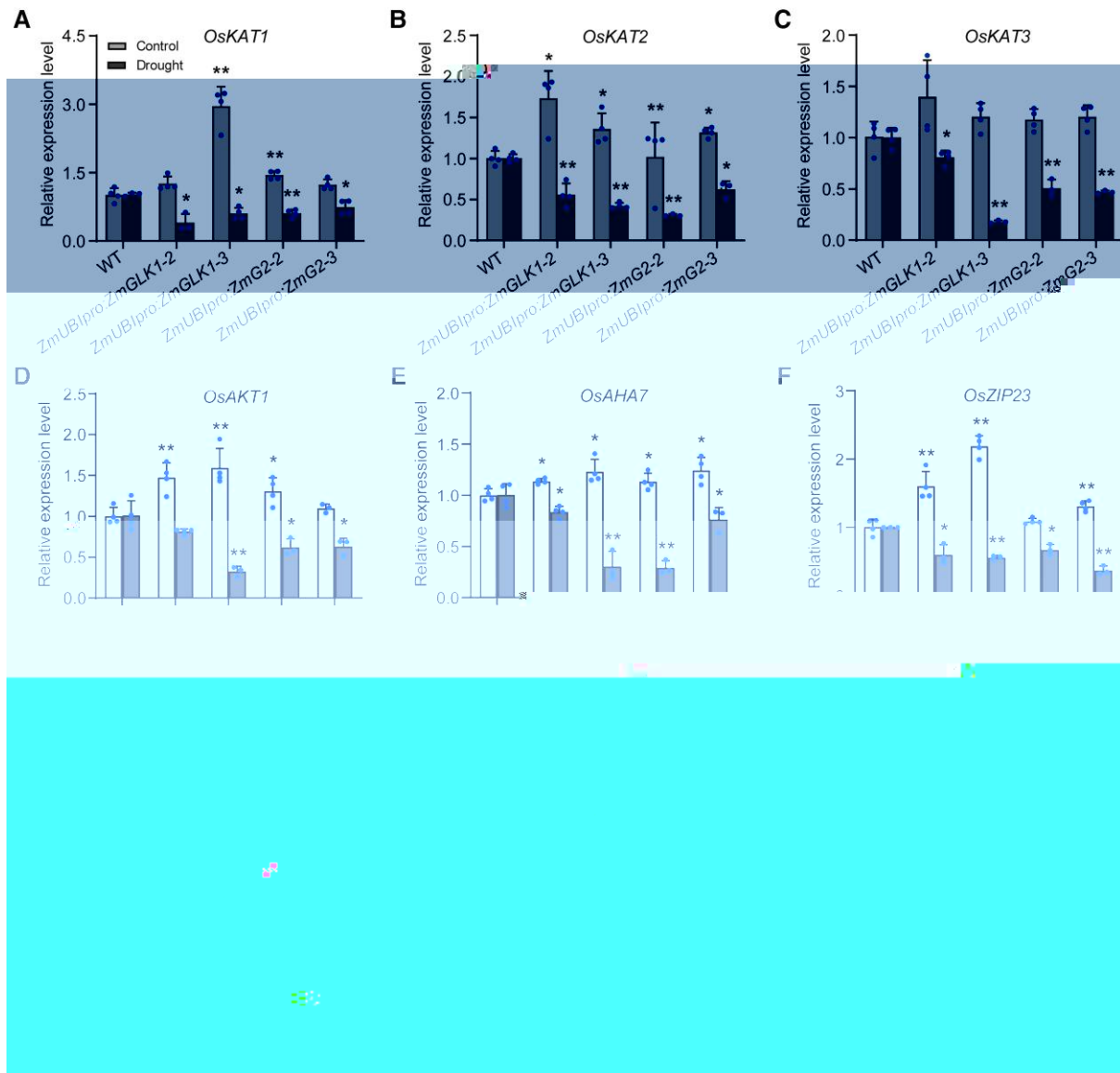


Figure 5. Relative expression levels of genes involved in stomatal movement and stomatal permeability in WT rice under normal conditions and 7 days of drought stress. Expression levels of (A) *OsKAT1*, (B) *OsKAT2*, (C) *OsKAT3*, (D) *OsAKT1*, (E) *OsAHA7*, (F) *OsZIP23*, (G) *OsGLK1*, and (H) *OsG2*. Gene expression levels were measured by RT-qPCR in the leaves of 3-week-old rice plants grown in soil under normal conditions or drought stress for 7 d. Data are presented as the mean \pm SD from 3 biological replicates. * < 0.05 , ** < 0.01 Student's *t*-test.

in plants overexpressing *OsGLK1* or *OsG2* (Fig. 6B; Supplemental Table S1). We noticed that upregulated DEGs were enriched in stomatal development and stomatal opening pathways in the DAP-seq analysis simultaneously. Therefore, these genes were identified as potential key genes of *ZmGLK1* and *ZmG2* in rice, including rice genes

OsKAT1, *OsKAT2*, *OsKAT3*, *OsAKT1*, *OsAHA7*, and *OsZIP23* (Fig. 7, A-D). The gene expression from RNA-seq analysis of these genes is prominently higher in WT rice plants (Fig. 7, E-H). Further reverse transcription

quantitative PCR (RT-qPCR) analysis verified that these genes were highly induced in WT rice under drought stress conditions (Fig. 7, I-L). These potential key genes may contribute to enhanced drought tolerance by enhancing stomatal movement when suffering from drought.

Discussion

GL TFs have been reported as some of the most important regulators of chloroplast genes and photosynthesis. They have been identified in Arabidopsis, tomato



Figure 6. Transcriptional profiles of WT, ZmUBlpro:ZmGLK1-3, and ZmUBlpro:ZmG2-3 in response to ABA treatment. **A)** PCA of gene expression profiles in WT, ZmUBlpro:ZmGLK1-3, and ZmUBlpro:ZmG2-3 in response to ABA treatment. **B)** Venn diagram showing the overlap of DEGs upregulated in RNA-seq and DAP-seq. **C, D)** GO enrichment analysis for DEGs upregulated in RNA-seq (C) and DAP-seq (D). Bubble size indicates the number of DEG counts in the corresponding GO category; bubble color corresponds to the $-\log_{10}$ of discovery rate [FDR] value, and the x-axis indicates the number of DEGs in each GO category.

Ross *et al.* 2001; Waters *et al.* 2009; Poelle *et al.* 2017). In rice, ectopic expression of maize *ZmGLK1* and *ZmGLK2* promotes pro- and stomatal gene expression, increases chloroplast and mitochondrial development in rice vascular sheath cells (Wang *et al.* 2017). A previous study by our lab has revealed that maize *ZmGLK1* overexpression in rice promotes stomatal density and increases the result of improved photosynthesis and reduced photo-inhibition under high light fluctuation conditions (Liu *et al.* 2020). In the present study, we uncovered the overexpression of maize *ZmGLK1* and *ZmGLK2* in rice enhanced drought tolerance by promoting stomatal closure. Specifically, transplants were grown under standard, salt, and drought conditions, we observed smaller stomatal density and higher stomatal density per square in rice plants overexpressing *ZmGLK1* or *ZmGLK2* compared to WT plants (Fig. 2, B and E). These results are consistent with our studies showing that *ZmGLK1* overexpression led

to increased stomatal conductance in field in rice (Liu *et al.* 2020), greenhouse in rice (Yeh *et al.* 2022), and Arabidopsis (Nishishiro *et al.* 2016). In contrast, under drought stress, the stomatal density of *ZmGLK1* or *ZmGLK2* overexpressing rice plants was reduced (Fig. 2, B and E), improving drought tolerance by preventing stomatal loss. Previous studies in rice have reported that high light density increases stomatal closure quickly, thus promoting resilience to drought stress (Cui *et al.* 2019; Cui *et al.* 2023). These prior results are consistent with those of the present study. Notably, differences in stomatal density between control and drought stressed plants were not observed for *ZmGLK1* or *ZmGLK2* overexpression, and were reduced by regulation of genes involved in stomatal movement, mainly in the + channels and the H⁺ATPase (Fig. 2, C and D; Fig. 5). Upregulation of + channels genes by *ZmGLK1* or *ZmGLK2* overexpression under normal conditions in line with previous study in Arabidopsis showing that *ZmGLK1* supports regulation of + channels genes and stomatal movement (Nishishiro *et al.* 2016). Thus, this rapid

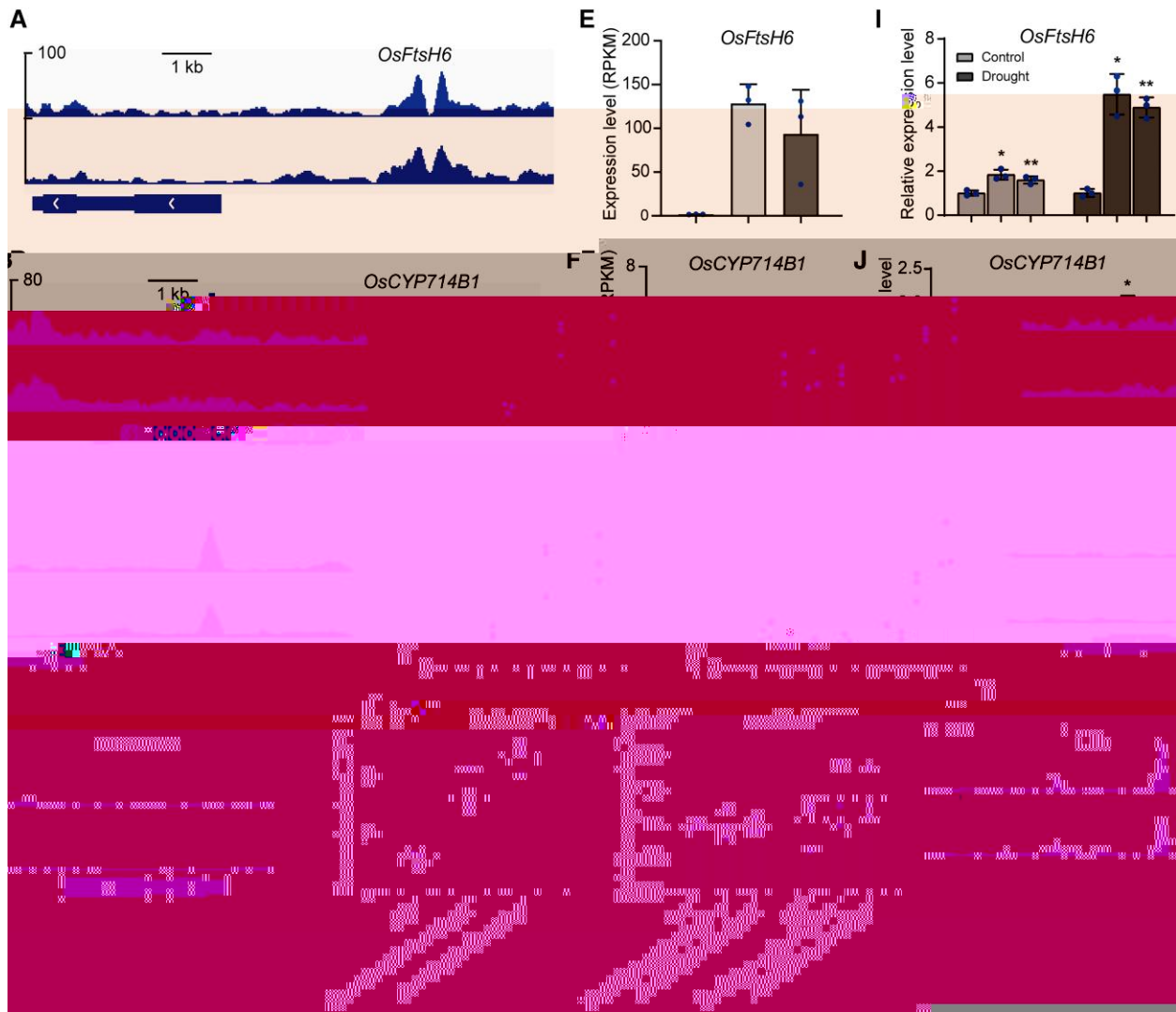


Figure 7. Purified ZmGL 1 and ZmGL 2 sequences in rice. **A to D)** DAP-seq tracks of ZmGL 1 and ZmGL 2 preferentially bound to the promoters of **A)** *OsFtsH6*, **B)** *OsCYP714B1*, **C)** *OsFtsH6*, and **D)** *OsCYP714B1*. **E to H)** Expression levels of **E)** *OsFtsH6*, **F)** *OsCYP714B1*, **G)** *OsFtsH6*, and **H)** *OsCYP714B1* in WT rice and rice overexpressing *OsFtsH6* or *OsCYP714B1*. **I to L)** Relative expression levels of **I)** *OsFtsH6*, **J)** *OsCYP714B1*, **K)** *OsFtsH6*, and **L)** *OsCYP714B1* in WT, *OsFtsH6*, and *OsCYP714B1* rice under control conditions and 7 d of drought stress. **D** Data are presented as the mean \pm SD from 3 biological replicates. * < 0.05, ** < 0.01. Student's *t*-test.

som closure of stomata in rice plants resulted directly from significant reduction in the expression levels of those genes under drought conditions.

Notably, the verified stomatal regulation of rice stomatal closure in response to ABA mediated by ABA signaling, supported by the exogenous application of ABA induction of stomatal closure in rice (Figure 4B). In rice, stomatal closure in WT rice (Figure 4B) is the effective of drought stress. Our findings are consistent with the previous study that stomatal closure requires the ABA signaling pathway (Cao and Sobrino 2020). Our results also implied that ZmGL 1 may function in the ABA biosynthesis pathway and is induced by the higher ABA accumulation (Supplemental Figure S5) by the bound and expression

of several key genes involved in ABA biosynthesis especially drought (Supplemental Figure S6). ABA biosynthesis starts with the epoxidation of xanthin, and xanthinophyll precursor therefore plays an important role in ABA biosynthesis. We previously discovered that ZmGL 1 increases levels of xanthinophylls, including xanthin and xanthin (Lee et al. 2020), which may lead to the improved ABA biosynthesis in rice. Moreover, study in Arabidopsis showed that ZmGL 1 directly controls the expression of *AtUGT74B1*, and ZmGL 1-WR Y40 may either directly regulate ABA signaling (Ahm et al. 2019) or possibly play a regulatory role of ZmGL 1 in the ABA signaling pathway. We also proposed that the Ca-like function conferred by ZmGL 1 is mentioned above may contribute to the rapid stomatal

closure. This has been demonstrated by models and experimental data for C_4 crops capable of more rapid stomatal closure compared to C_3 crops in response to the decrease in leaf water potential and use efficiency WUE (McAusland [et al.](#) 2016; Wang [et al.](#) 2011; Oke [et al.](#) 2011). Notably, previous studies have demonstrated that suberisation and closure in ferns is associated with reduced responsiveness to ABA and sugars compared to gymnosperms (Lim [et al.](#) 2019; Cabeldo-Sobrinho [et al.](#) 2011). In the present study, the relationship between the cell wall and suberisation in fern species contributes to the stomatal movement (Chen [et al.](#) 2017).

Spore germination and growth

Red yeast were detached from control or drought stressed 3- to 4-week-old rice seedlings grown in pots under normal conditions or drought stress for 7 d. Samples were flamed in liquid nitrogen and ground to powder, and then RNA was extracted using TRIzol reagent. RNA purification and quantification were performed using a NanoDrop 2000 spectrophotometer. ThermoFisher Scientific, USA. First-strand cDNA synthesis was performed using the ReverTra-First-Strand cDNA Synthesis Kit (ThermoFisher Scientific, USA). RT-qPCR was performed using the SYBR Green mix (TOYOBO) on the AB Q9700 6 Flex instrument. Applied Biosystems, USA. Relative mRNA levels were calculated using the $2^{-\Delta\Delta CT}$ method (Livak and Schmittgen, 2001) with 3 biological replicates for each treatment using the *act1* gene as a control. Primers are listed in Supplemental Table S1.

RNA-seq analysis

The uppermost fully expanded leaves of control and drought stressed rice seedlings were detached and flamed in liquid nitrogen. Ground samples (100 mg each) were extracted in the lysis solution containing a protein inhibitor cocktail (Roche) and 500 mg/ml chloroform. Samples were centrifuged, and the resulting supernatant was extracted again. The combined extracts were purified on a C₁₈ silica column and dried in a rotary evaporator. First-strand cDNA synthesis was performed using the SuperScript III RT kit (Life Technologies, USA). ABA was quantified using HPLC-MS/MS system as described by Liu et al. (2012).

Exogenous ABA treatment

Four-week-old rice seedlings grown in pots were sprayed with 100 μM ABA solution containing 0.5% [v/v] Tween-20 to the surface of the leaves. The volume of ABA solution applied was constant between seedlings. A 5-hr drought treatment was exchanged with the same volume of water and described above.

RNA extraction and RT-qPCR

The uppermost fully expanded leaves were harvested from 3- to 4-week-old rice seedlings grown in pots under normal conditions or drought stress for 7 d. Samples were flamed in liquid nitrogen and ground to powder, and then RNA was extracted using TRIzol reagent. RNA purification and quantification were performed using a NanoDrop 2000 spectrophotometer. ThermoFisher Scientific, USA. First-strand cDNA synthesis was performed using the ReverTra-First-Strand cDNA Synthesis Kit (ThermoFisher Scientific, USA). RT-qPCR was performed using the SYBR Green mix (TOYOBO) on the AB Q9700 6 Flex instrument. Applied Biosystems, USA. Relative mRNA levels were calculated using the $2^{-\Delta\Delta CT}$ method (Livak and Schmittgen, 2001) with 3 biological replicates for each treatment using the *act1* gene as a control. Primers are listed in Supplemental Table S1.

RNA-seq analysis

After exogenous ABA treatment, leaves were collected from 4-week-old rice seedlings grown in pots. Total RNA was extracted using TRIzol reagent and then RNA purification was performed using the RNeasy RiboZero RRNA Removal Kit (Qiagen, USA). RNA-seq libraries were constructed from WT, *act1*, and *act2* rice plants using the TruSeq Sampled mRNA LT Sample Prep Kit (Illumina, USA) with 3 biological replicates per line. The resulting libraries were sequenced on the Illumina HiSeq X Ten sequencing platform. After removing the adaptor sequences and low-quality reads, clean reads were mapped to the rice cv. Nipponbare reference genome using HISAT2 (Mao et al., 2015) and Bowtie2 (Langmead and Salzberg, 2009). Gene expression levels were calculated as reads per kilobase of transcript per million mapped reads (RPKM) using cuffdiff. DEGs were identified using the DESeq2 R package. The thresholds for differential expression were $p < 0.05$ and $|\log_2(\text{fold change})| > 1$.

DAP-seq analysis

The full-length coding sequences of *act1* and *act2* were amplified from cDNA of the mRNA accessions B73. Each sequence was recombined into the pY-HALO vector using LR Clonase II Inverse. The pY-HALO-ZmGL1 and pY-HALO-ZmG promoters were generated using 500 bp each of the pY-HALO-ZmGL1 and pY-HALO-ZmG promoters (Bartlett et al., 2010).

conserved sequences $P < 0.05$. Figures were generated using GraphPad Prism 9.0 and Adobe Illustrator CS3.

Accession numbers

RNA-sequence data were deposited in the NCB BioProject database under accession number PRJNA1018861 for RNA-seq and PRJNA1019016 for DAP-seq. The sequence data from this study can be found in the GenBank/EMBL database under the following accession numbers: GenBank ID: AF318580 and GenBank ID: AF318579.

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Author contributions

W.Z. and J.L. conceived and designed the experiment. J.L., J.L., S.W., Y.G., and R.G. performed most of the experiment. Z.L. and H.P. performed the DAP-seq experiment. W. critically commented and edited the manuscript. W. primarily prepared by J.L., J.L., and W.Z. All authors discussed and commented on the manuscript.

Supplemental data

The following materials are available in the online version of this article.

Supplemental Figure S1. Enhanced tolerance of rice plants to drought stress induced by 20% PEG 6000.

Supplemental Figure S2. Overexpression of *ZmLs* in rice led to decreased stomatal conductance and photosynthesis parameters in response to drought.

Supplemental Figure S3. Dynamic changes of soil carbon during the drought stress in the rearing house experiment.

Supplemental Figure S4. Genome-wide summary of the regulatory network of *ZmGL1* and *ZmGtb* based on DAP-seq data.

Supplemental Figure S5. Changes in endogenous ABA content in WT, *ZmLs* overexpressing rice lines under normal conditions and 7 d of drought stress.

Supplemental Figure S6. Relative expression levels of ABA biosynthesis genes in the leaves of WT, *ZmLs* overexpressing rice plants under normal conditions and 7 d of drought stress.

Supplemental Table S1. Relative changes of gene expression levels of 59 overlapped genes from RNA-seq and DAP-seq analyses.

Supplemental Table S2. Primers used for RT-qPCR.

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The authors declare that they have no conflict of interest.

Data availability

The data underlying this article are available in the public domain online supplementary material.

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